

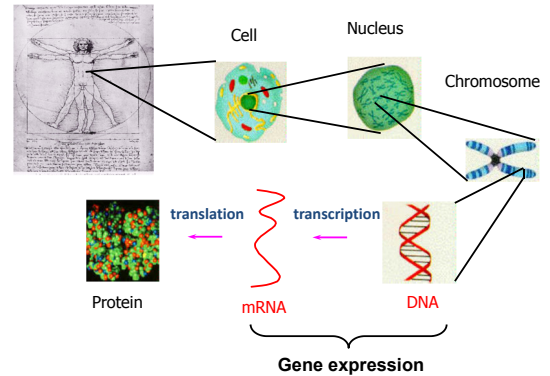
Lecture 16:
Microarray data analysis: introduction

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<http://www.cs.otago.ac.nz/cosc348/>

1

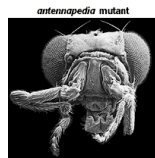
Microarrays measure gene expression



2



Gene Expression

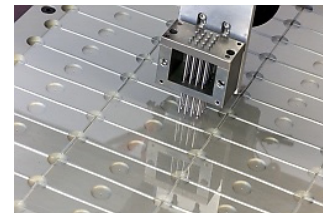


- Cells are different because of **differential gene expression** (i.e. in different body organs different genes are expressed).
- Gene is expressed by transcribing DNA into **many copies** of mRNA.
- mRNAs are then **translated** into protein molecules.
- **Microarrays measure the level of mRNA (i.e. concentration of mRNA), and thus the level of gene expression.**

3

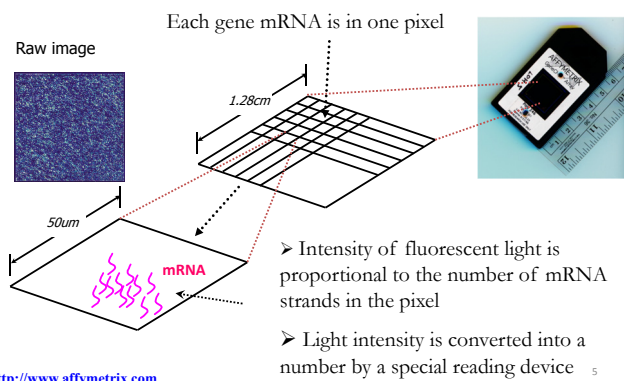
Principle of microarrays

- mRNA levels are proportional to the rate (or level) of gene expression (*how many copies of mRNA are produced by a gene*)
- mRNA is isolated from cells and labeled with a fluorescent dye.
- **Level of mRNA is proportional to intensity of fluorescent light emission, which is measured.**



4

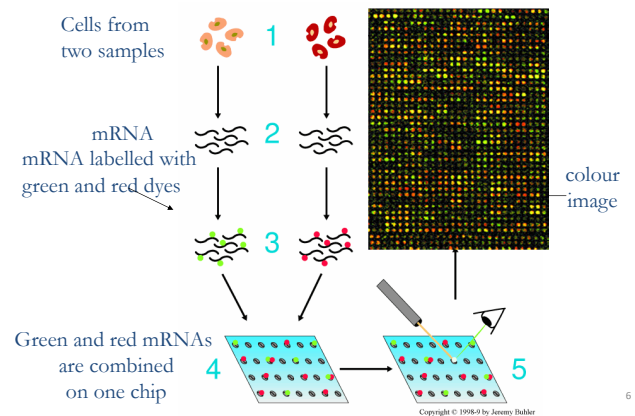
Affymetrix microarrays



5

<http://www.affymetrix.com>

Spotted or cDNA microarrays



6

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Image analysis in spotted arrays

- The 2 fluorescence images are overlaid, with the colours combined (red and green make yellow in the RGB scheme).
- Yellow spots** indicate genes which were equally expressing in both conditions/ cell groups (i.e. **both genotypes**)
- Green spots** indicate genes which were only expressing in the control condition (e.g., **wild-type genotype**).
- Red spots** indicate genes which were only expressing in the treatment condition (e.g., **mutant genotype**).

7

Fold changes in spotted arrays

- Differential expression at a spot is often reported as a fold change:

$$\text{Fold change} = \frac{\text{red intensity}}{\text{green intensity}}$$

- In spotted arrays too, the light intensity is converted into a numerical value (fold change) by a special equipment.

- Often \log_2 scale is used:

$$\begin{aligned} \log_2(\text{Fold change}) &= \log_2\left(\frac{\text{red intensity}}{\text{green intensity}}\right) = \\ &= \log_2(\text{red intensity}) - \log_2(\text{green intensity}) \end{aligned}$$

8

Microarray data – N x M matrix of numbers

M samples / subjects

ID	WT_1_R	WT_2_R	WT_3_R	WT_4_R	KO_1_R	KO_2_R	KO_3_R	KO_4_R
93173_at	242.3	240.1	292.9	216.3	180.1	172.6	147.3	152.4
101937_s_at	316.7	346.7	438.3	228.5	133.7	201.3	253.3	287.4
104272_s_at	286.2	351.9	354.6	339.1	180.6	432.7	210.2	53.6
98590_at	1,066	748.8	1,011.4	607.7	584.5	791.8	355.8	530
102425_at	264.7	241.4	450	134.3	138.3	242	212.6	125.4
96608_at	1,979.8	1,913.2	2,367	1,616	1,270.5	1,191.6	1,401.2	1,330.9
94407_at	339.3	360.4	283	309.1	236.9	329.3	196.8	89.4
161149_f_at	1,947.7	1,179.4	1,708	1,251	1,297.1	594.3	1,070.5	1,055.8
100144_at	4,821.6	3,639.6	4,415.5	3,846	3,288.5	2,438.5	2,799	2,537.4
95134_at	498.6	853.1	881.2	582.8	255.1	859.3	288.7	457.8
96921_at	746.1	410.6	858.8	667.4	534.8	444	475.4	320.3
94689_at	534	438	456.2	555.2	466.6	404.3	295.2	146.4
160268_at	737.7	1,099.2	1,138.4	978.8	806.5	978	587.8	245.3
96180_at	609.5	516.9	540.1	312.8	344.8	191.8	427.9	347.1
92618_at	4,888.8	4,234.2	4,703.7	2,994.9	4,093.1	2,938.9	2,150.2	1,969.2
95203_f_at	111.8	186.8	112.9	158.1	100.8	67	119.9	90.6
102574_at	171.3	81.7	230.9	123.3	107.9	50.6	112.3	132.4
160966_at	221.2	310	454.3	242.5	238	196.2	330.7	50.8
160827_at	294.5	341.1	360.4	170.3	231.6	289.4	196.4	58.1
104116_at	1,836.3	829.3	1,258.7	1,561	722.3	810.4	943.9	1,172.1
95434_at	1,207.8	1,294.8	1,314.6	1,513.8	878.2	773.9	715.8	1,181.5

WT = wild-type (i.e. all genes present in the genome);
KO = gene knock-out (one gene is removed/silenced)

9

Matrix description

- Microarray data can be viewed as an N x M matrix:

– Each of the N rows represents a gene

– Each of the M columns represents a sample (e.g., patient, animal, etc.)

– Each matrix pixel represents the *expression level* of a gene. It can be either an absolute value (e.g. Affymetrix GeneChip) or a relative expression ratio (e.g. spotted microarrays).

– A row is referred to as the “*expression profile of the gene*”.

– A column is referred to as the “*expression profile of the sample*”.

10

Microarray data mining challenges

- too few records (samples, animals, patients), usually < 100
- too many columns (genes), usually $1,000 < \# < 10,000$
- for exploration, a large set of all relevant genes is desired
- for diagnostics or identification of therapeutic targets, the smallest set of genes is needed
- model needs to be explainable to biologists

11

Differential gene expression analysis

- The Experiment measures gene expression in rats:
 - Two groups: (WT: wild-type rat, KO: gene knock-out rat)
 - Question: Which genes are affected by the treatment? How significant is the effect? **We compare each pair of genes.**

ID	WT_1_R	WT_2_R	WT_3_R	WT_4_R	KO_1_R	KO_2_R	KO_3_R	KO_4_R
93173_at	242.3	240.1	292.9	216.3	180.1	172.6	147.3	152.4
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12

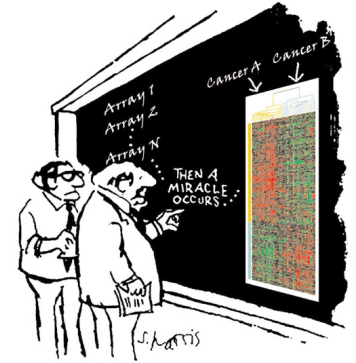
Experiments and questions

- Two-condition comparison against some form of control:
 - Gene knock-out against wild-type (KO vs. WT)
 - Subjects with a disease vs. healthy subjects
 - Treated subjects vs. untreated subjects
 - Etc
- Question of interest in these experiments are:
 - which genes are influenced by the missing gene?
 - which genes are responsible for the disease?
 - which genes are influenced by the administered drug?
 - Etc.

13

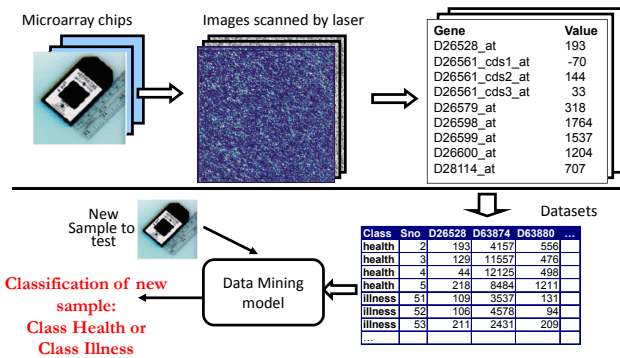
Goals of a Microarray Experiment

- Find the genes** that change expression between experimental and control samples
- Find patterns:** Groups of biologically related genes that change expression together.
- Classify new samples** based on a gene expression profile.



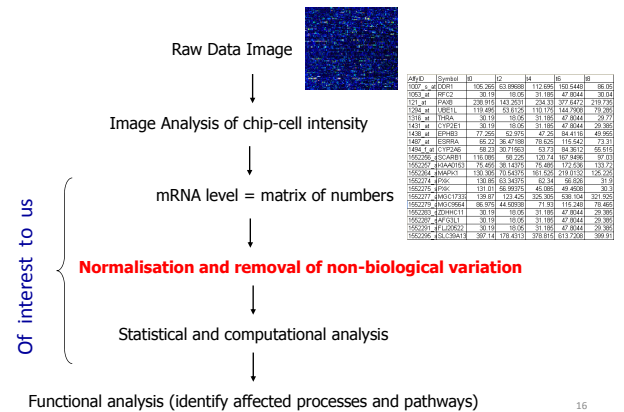
14

Goal: develop a model to classify a new sample



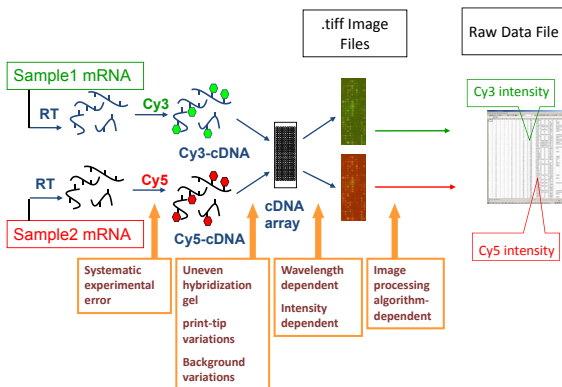
15

Steps in microarray data analysis



16

Microarray data are very noisy



17

Removal of noise: thresholding & filtering

- Thresholding:** Removing bad intensity spots is an important process of quality control. For example, the scanner has a measurement limit below which intensity values cannot be trusted. Values below the cut-off point are usually removed (filtered) from the data because they are likely to be artifacts.
 - Typically, the lowest intensity value of reliable data is 100–200 for Affymetrix data and 100–1000 for cDNA microarray data.
- Filtering:** remove genes with insufficient variation between two conditions:
 - e.g. $\text{MaximumValue} - \text{MinimumValue} < \Delta$ (usually 500)
 - $\text{MaximumValue} / \text{MinimumValue} < r$ (usually 5)
- Input for further processing is a matrix of numbers.

18

Normalisation

- Gene expressions can differ by an order of magnitude.
- Normalisation is needed for gene selection, clustering and classification models.
- **Normalisation:** mathematical transformation of values of gene expression from the interval $[m_{\min}, m_{\max}] \rightarrow [m'_{\min}, m'_{\max}]$, either
 - Linearly
 - Logarithmically
 - to Mean = 0, Std. Dev = 1
 - other
- **Whatever the method: normalise each gene row separately!**

19

Linear normalisation

- **Linear Normalisation:** Let m' be the new normalised value of gene expression / mRNA level:

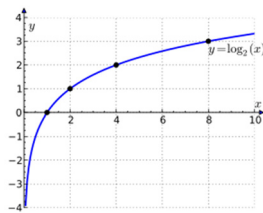
$$m' = \frac{m - m_{\min}}{m_{\max} - m_{\min}}$$

- This equation transforms values of gene expressions from the interval $[m_{\min}, m_{\max}] \rightarrow [0, 1]$ **uniformly**.
 - When $m = m_{\min}$, then $m' = 0$
 - When $m = m_{\max}$, then $m' = 1$

20

Logarithmic normalisation

- If the data have a huge value span like from 10^2 to 10^4 , then it's more suitable to use a logarithm, e.g. $m' = \log m$. (Either \log_{10} or \log_2).
- This equation transforms values of gene expressions from the interval $[m_{\min}, m_{\max}] \rightarrow [m'_{\min}, m'_{\max}]$ **nonuniformly**.



Other equations for normalisation:

<http://people.revoledu.com/kardi/tutorial/Similarity/Normalization.html>

21

What's next after normalisation:

- Gene Selection
 - find genes, which would be the best predictors (of disease, treatment outcome, etc.)
- Clustering (Unsupervised, no class labels)
 - Exploration and finding patterns
 - find new biological classes of genes / refine existing ones
- Classification (Supervised, needs class labels)
 - identify disease and its genetic profile
 - predict outcome / select best treatment
- Functional / ontology analysis

22

Potential applications of microarrays

- Biological and medical discovery
 - discovery of putative functions of genes
 - finding and refining biological pathways
 - new and better molecular diagnostics / “personalised” medicine
 - appropriate treatment for genetic signatures
 - potential genetic targets for new therapies

23